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COLLABORATION

Animal data reliance bias in peer review: a barrier to scientific progress

Marcia Triunfol, PhD

Biomedical Science Advisor, Research & Toxicology

mtriunfol@hsi.org



<https://app.sli.do/event/4zlwcyln>

Early definition of publication bias

Publication Bias: The Problem That Won't Go Away

KAY DICKERSIN

*Department of Epidemiology and Preventive Medicine
University of Maryland School of Medicine
Howard Hall
660 West Redwood Street
Baltimore, Maryland 21201*


YUAN-I MIN

*Department of Epidemiology
The Johns Hopkins University
School of Hygiene and Public Health
615 North Wolfe Street
Baltimore, Maryland 21205*

[Publication bias: the problem that won't go away.](#) Dickersin K, Min YI. Ann N Y Acad Sci. 1993 Dec 31;703:135-46; discussion 146-8. doi: 10.1111/j.1749-6632.1993.tb26343.x.PMID: 8192291

Catalogue of publication bias

Catalogue of Bias



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Publication bias

When the likelihood of a study being published is affected by the findings of the study.

Background

Dickersin & Min define publication bias as the failure to publish the results of a study "on the basis of the direction or strength of the study findings." This non-publication introduces a bias which impacts the ability to accurately synthesize and describe the evidence in a given area. Publication bias is a type of reporting bias and closely related to dissemination bias, although dissemination bias generally applies to all forms of results dissemination, not simply journal publications. A variety of distinct biases are often grouped into the overall definition of publication bias.

There are a number of reasons for publication bias identified in the literature. Research has shown causes of publication bias ranging from trialist motivation, past experience, and competing commitments; perceived or real lack of interest in results from editors, reviewers or other colleagues; or conflicts of interest that would lead to the suppression of results not aligned with a specific agenda.

Cite as

Catalogue of bias collaboration, Devito N, Goldacre B. **Publication bias**. In Catalogue Of Bias. 2019. <https://catalogofbias.org/biases/publicationbias/>

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Catalogue of bias collaboration, Devito N, Goldacre B. **Publication bias**. In Catalogue Of Bias. 2019. <https://catalogofbias.org>

Seminal work on publication bias

J Chron Dis Vol. 32, pp. 51 to 63
Pergamon Press Ltd 1979. Printed in Great Britain

BIAS IN ANALYTIC RESEARCH

DAVID L. SACKETT

INTRODUCTION

CASE-CONTROL studies are highly attractive. They can be executed quickly and at low cost, even when the disorders of interest are rare. Furthermore, the execution of pilot case-control studies is becoming automated; strategies have been devised for the 'computer scanning' of large files of hospital admission diagnoses and prior drug exposures, with more detailed analyses carried out in the same data set on an *ad hoc* basis [1]. As evidence of their growing popularity, when one original article was randomly selected from each issue of **The New England Journal of Medicine**, **The Lancet**, and the **Journal of the American Medical Association** for the years, 1956, 1966 and 1976, the proportion reporting case-control analytic studies increased fourfold over these two decades (2–8%) whereas the proportion reporting cohort analytic studies fell by half (30–15%); incidentally, a general trend toward fewer study subjects but more study authors was also noted [2].

Early resistance to accept the failure of mice models



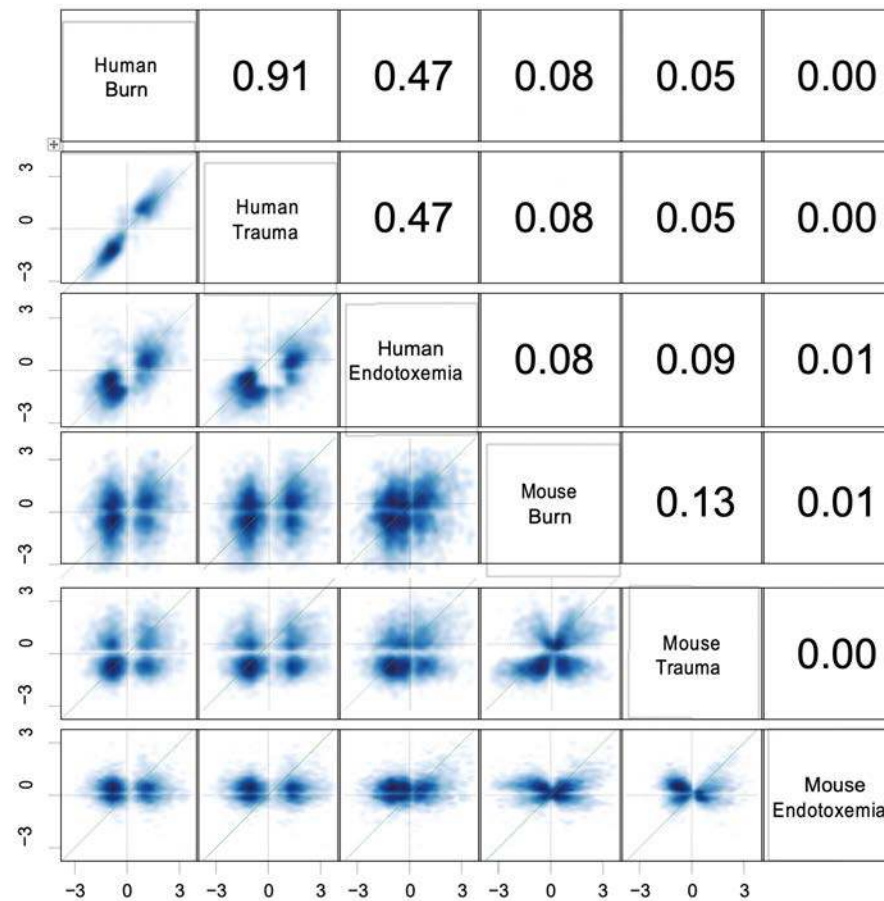
“They [the reviewers] were so used to doing mouse studies that they thought that was how you validate things.”

Genomic responses in mouse models poorly mimic human inflammatory diseases

Junhee Seok^{a,1}, H. Shaw Warren^{b,1}, Alex G. Cuenca^{c,1}, Michael N. Mindrinos^a, Henry V. Baker^c, Weihong Xu^a, Daniel R. Richards^d, Grace P. McDonald-Smith^e, Hong Gao^a, Laura Hennessy^f, Celeste C. Finnerty^g, Cecilia M. López^c, Shari Honari^f, Ernest E. Moore^h, Joseph P. Mineiⁱ, Joseph Cuschieri^j, Paul E. Bankey^k, Jeffrey L. Johnson^h, Jason Sperry^l, Avery B. Nathens^m, Timothy R. Billiar^l, Michael A. Westⁿ, Marc G. Jeschke^o, Matthew B. Klein^j, Richard L. Gamelli^p, Nicole S. Gibran^j, Bernard H. Brownstein^q, Carol Miller-Graziano^k, Steve E. Calvano^r, Philip H. Mason^e, J. Perren Cobb^s, Laurence G. Rahme^t, Stephen F. Lowry^{r,2}, Ronald V. Maier^j, Lyle L. Moldawer^c, David N. Herndon^g, Ronald W. Davis^{a,3}, Wenzhong Xiao^{a,t,3}, Ronald G. Tompkins^{t,3}, and the Inflammation and Host Response to Injury, Large Scale Collaborative Research Program⁴

^aStanford Genome Technology Center, Stanford University, Palo Alto, CA 94305; Departments of ^bPediatrics and Medicine, ^cAnesthesiology and Critical Care Medicine, and ^dSurgery, Massachusetts General Hospital, Harvard Medical School, Boston, MA 02114; ^eDepartment of Surgery, University of Florida College of Medicine, Gainesville, FL 32610; ^fIngenuity Inc., Redwood City, CA 94063; ^gDepartment of Surgery, Massachusetts General Hospital, Boston, MA 02114; ^hDepartment of Surgery, Harborview Medical Center, Seattle, WA 98195; ⁱShriners Hospitals for Children and Department of Surgery, University of Texas Medical Branch, Galveston, TX 77550-1220; ^jDepartment of Surgery, University of Colorado Anschutz Medical Campus, Denver, CO 80045; ^kDepartment of Surgery, Parkland Memorial Hospital, University of Texas, Southwestern Medical Center, Dallas, TX 75390; ^lDepartment of Surgery, Harborview Medical Center, University of Washington School of Medicine, Seattle, WA 98195; ^mDepartment of Surgery, University of Rochester School of Medicine, Rochester, NY 14642; ⁿDepartment of Surgery, University of Pittsburgh Medical Center Presbyterian University Hospital, University of Pittsburgh, PA 15213; ^oDepartment of Surgery, St. Michael's Hospital, University of Toronto, Toronto, ON, Canada M5B 1W8; ^pDepartment of Surgery, San Francisco General Hospital, University of California, San Francisco, CA 94143; ^qDivision of Plastic and Reconstructive Surgery, Department of Surgery, University of Toronto, Toronto, ON, Canada M4N 3M5; ^rDepartment of Surgery, Stritch School of Medicine, Loyola University, Chicago, IL 60153; ^sDepartment of Anesthesiology, Washington University, School of Medicine, St. Louis, MO 63110; and ^tDepartment of Surgery, University of Medicine and Dentistry of New Jersey-Robert Wood Johnson Medical School, New Brunswick, NJ 08903

Contributed by Ronald W. Davis, January 7, 2013 (sent for review December 6, 2012)



Junhee Seok, H. Shaw Warren, Alex GC, Michael NM, Henry VB, Xu W, et al. Genomic responses in mouse models poorly mimic human inflammatory diseases. Proc Natl Acad Sci U S A. 2013;110: 3507–3512.

Evidence of animal data reliance bias



bioRxiv posts many COVID19-related papers. A reminder: they have not been formally peer-reviewed and should not guide health-related behavior or be reported in the press as conclusive.

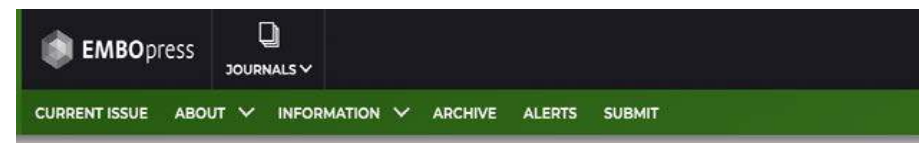
New Results

Long-term expanding human airway organoids for disease modelling

Norman Sachs, Dominique D. Zomer-van Ommen, Angelos Papaspyropoulos, Inha Heo, Lena Böttinger, Dymph Klay, Fleur Weeber, Guizela Huelsz-Prince, Nino Jakobachvili, Marco C. Viveen, Anna Lyubimova, Luc Teeven, Sepideh Derakhshan, Jeroen Korving, Harry Begthel, Kuldeep Kumawat, Emilio Ramos, Matthijs F.M. van Oosterhout, Eduardo P. Olimpio, Joep de Ligt, Krijn K. Dijkstra, Egbert F. Smit, Maarten van der Linden, Emile E. Voest, Coline H.M. van Moorsel, Cornelis K. van der Ent, Edwin Cuppen, Alexander van Oudenaarden, Frank E. Coenjaerts, Linde Meyaard, Louis J. Bont, Peter J. Peters, Sander J. Tans, Jeroen S. van Zon, Sylvia F. Boj, Robert G. Vries, Jeffrey M. Beekman, Hans Clevers

doi: <https://doi.org/10.1101/318444>

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Resource | 14 January 2019 | [OPEN ACCESS](#)

[TRANSPARENT PROCESS](#)

Long-term expanding human airway organoids for disease modeling


Norman Sachs, Angelos Papaspyropoulos, Dominique D. Zomer-van Ommen, Inha Heo, Lena Böttinger, Dymph Klay, Fleur Weeber, Guizela Huelsz-Prince, Nino Jakobachvili, Gimano D. Amatngalim, Joep de Ligt, Arne van Hoeck, Natalie Proost, Marco C. Viveen, Anna Lyubimova, Luc Teeven, Sepideh Derakhshan, Jeroen Korving, Harry Begthel, Johanna F. Dekkers, Kuldeep Kumawat, Emilio Ramos, Matthijs F.M. van Oosterhout, G. Johan Offerhaus, [...] Hans Clevers

[Author Information](#)

EMBO J (2019) 38: e100300 | <https://doi.org/10.15252/embj.2018100300>

See also: [M Paschini & CF Kim](#) (February 2019)

“This is a popular paper that was very difficult to publish. We had to try three or four different journals, until we had it accepted. I don’t recall exactly at what point we decided to add the animal experiments, but I believe it was done because of a request by an editor or referee from one of these journals,” says Prof. Dr Clevers.



The screenshot shows the EMBOpress website interface. At the top, there is a dark header with the EMBOpress logo and a 'JOURNALS' dropdown menu. Below this is a green navigation bar with links for 'CURRENT ISSUE', 'ABOUT', 'INFORMATION', 'ARCHIVE', 'ALERTS', and 'SUBMIT'. The main content area has a light background. It features a 'Resource' label, the date '14 January 2019', an 'OPEN ACCESS' badge, and a 'TRANSPARENT PROCESS' icon. The article title is 'Long-term expanding human airway organoids for disease modeling'. The authors listed are Norman Sachs, Angelos Papaspyropoulos, Dominique D Zomer-van Ommen, Inha Heo, Lena Böttinger, Dymph Klay, Fleur Weeber, Guizela Huelsz-Prince, Nino Iakobachvili, Gimano D Amatngalim, Joep de Ligt, Arne van Hoeck, Natalie Proost, Marco C Viveen, Anna Lyubimova, Luc Teeven, Sepideh Derakhshan, Jeroen Korving, Harry Begthel, Johanna F Dekkers, Kuldeep Kumawat, Emilio Ramos, Matthijs FM van Oosterhout, G Johan Offerhaus, and Hans Clevers. There is an 'Author Information' link. The article is cited as 'EMBO J (2019) 38: e100300' with a DOI link. A 'See also' section mentions 'M Paschini & CF Kim (February 2019)'.

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Long-term expanding human airway organoids for disease modeling

Norman Sachs, Angelos Papaspyropoulos, Dominique D Zomer-van Ommen, Inha Heo, Lena Böttinger, Dymph Klay, Fleur Weeber, Guizela Huelsz-Prince, Nino Iakobachvili, Gimano D Amatngalim, Joep de Ligt, Arne van Hoeck, Natalie Proost, Marco C Viveen, Anna Lyubimova, Luc Teeven, Sepideh Derakhshan, Jeroen Korving, Harry Begthel, Johanna F Dekkers, Kuldeep Kumawat, Emilio Ramos, Matthijs FM van Oosterhout, G Johan Offerhaus, [...] Hans Clevers

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Scientists speak out

PROGRESS REPORT

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Is it Time for Reviewer 3 to Request Human Organ Chip Experiments Instead of Animal Validation Studies?

Donald E. Ingber

For the past century, experimental data obtained from animal studies have been required by reviewers of scientific articles and grant applications to validate the physiological relevance of in vitro results. At the same time, pharmaceutical researchers and regulatory agencies recognize that results from preclinical animal models frequently fail to predict drug responses in humans. This *Progress Report* reviews recent advances in human organ-on-a-chip (Organ Chip) microfluidic culture technology, both with single Organ Chips and fluidically coupled human "Body-on-Chips" platforms, which demonstrate their ability to recapitulate human physiology and disease states, as well as human patient responses to clinically relevant drug pharmacokinetic exposures, with higher fidelity than other in vitro models or animal studies. These findings raise the question of whether continuing to require results of animal testing for publication or grant funding still makes scientific or ethical sense, and if more physiologically relevant human Organ Chip models might better serve this purpose. This issue is addressed in this article in context of the history of the field, and advantages and disadvantages of Organ Chip approaches versus animal models are discussed that should be considered by the wider research community.

their premise is that the results they generate in their cell cultures will translate to humans. Because in vitro studies generally lack the natural three dimensional (3D) context, vascular flow, and physico-chemical microenvironment of living tissues and organs, as well as the multi-organ physiology of whole organisms, many question the clinical relevance of findings obtained with these simplified models. For this reason, most researchers who submit a grant application or publication based on in vitro findings commonly expect to find at least one reviewer (the classic exasperating "Reviewer 3") who demands that additional animal experiments be carried out to validate their findings before the work could be acceptable for publication or funding. This article seeks to provoke a conversation in the scientific community by asking two simple questions: does this make sense, and if not, is there a better alternative? I address these questions by reviewing recent progress that has been made using organoids and engineered microphysiological systems (MPS) with a focus on microfluidic organ-on-a-chip (Organ Chip) culture technologies.

1. Introduction

Ingber DE. Is it Time for Reviewer 3 to Request Human Organ Chip Experiments Instead of Animal Validation Studies? Adv Sci (Weinh). 2020 Oct 12;7(22):2002030.



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Physicians
Committee
for Responsible Medicine

Survey to Assess Journal and Reviewer Requests for Evidence in Animals



Q: During manuscript submission peer review, how many times have you been asked for animal experimental data to be added to a study that otherwise had no animal-based experiments? Did you feel the requested additional animal-based experiments were justified?

*“Sometimes the reviewers identify critical gaps in knowledge, these are valuable peer reviews. **Other times it seems like they ask for animals out of habit. We refuse.** This is even more difficult and hard to deal with when it comes to grant reviews.”*

*“The study was about heterogeneity of cancer cells from human tissue samples.
It was irrelevant to do an experiment on mice.”*

*“Referees ask for animal experiments because it is customary to do so in the field of biophysics, toxicology not because it is necessary. **Many researchers are unaware about the potential of in vitro, in silico methods and human based models.**”*

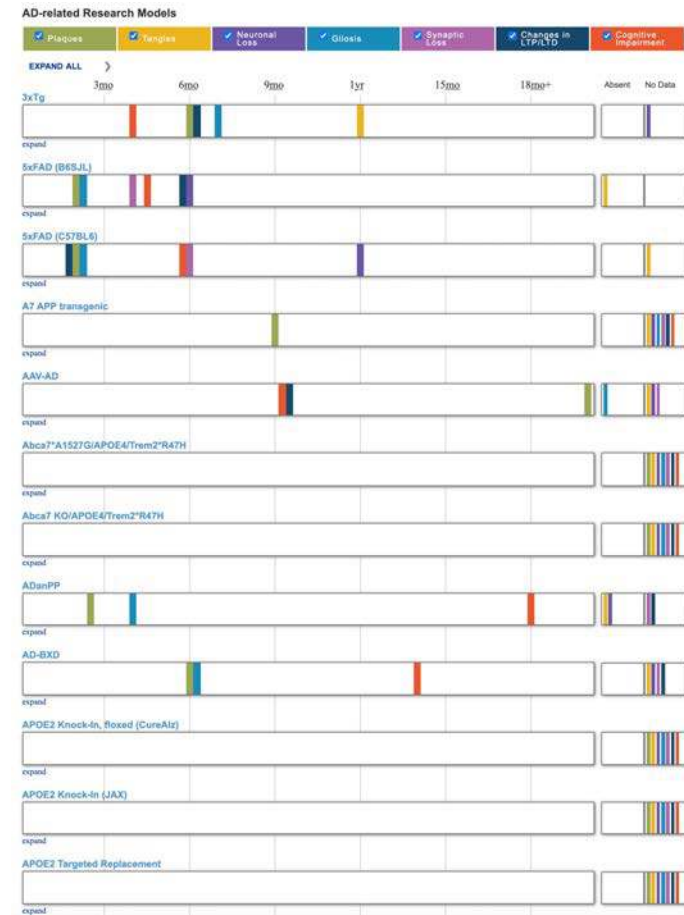
*“The need for validation of human organoid data with animal studies...
just because journal reviewers were used to this.”*

“They wanted human in vitro data to be “validated” against an animal model.”

Possible reasons behind animal data reliance bias

- Lack of understanding of how advanced human-based technologies work
- Status quo
- Journal editorial policy
- Scientific justification
- Regulatory requirement
- Avoiding sunk costs and the bygones principle

More than 200 rodent models of Alzheimer's Disease



How to prevent animal data reliance bias?

- Explore the scientific, ethical and economic advantages of advanced animal-free models
- Emphasize NAMs and non-animal research design in academic curricula and continuing education programs
- Recommend amendments to journal policies
- Discuss (challenge) scientific justifications (when claimed to exist) and regulation

Toward a new best practice

- Editors of scientific journals should **scrutinize** reviewer feedback requesting animal data to be supplied or generated *de novo* to validate or complement findings from advanced non-animal approaches as a condition for publication and **require a high level of justification** for such requests
- Journals should commit to **publicly disclose** when such requests are made



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Marcia Triunfol, mtriunfol@hsi.org

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